# SYNTHESIS OF 5-SUBSTITUTED-3-PHENACYLOXIME-1,2,4-OXADIAZOLES FROM ACETYLATED OR BENZOYLATED BENZOYLTHIACETAMIDE WITH HYDROXYLAMINE HYDROCHLORIDE

## SPECTRAL EVIDENCE FOR A TETRAHEDRAL CARBINOLAMINE INTERMEDIATE

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Abstract—Acetylated and benzoylated benzoylthiacetamide 1a, b reacts with hydroxylamine hydrochloride in refluxing ethanol to yield 3 - acetylamino - and 3 - benzoylamino - 5 - phenyl - isoxazoles 2a, b, while 3 - phenacyloxime - 5 - substituted - 1,2,4 - oxadiazoles 4a, b are formed in the presence of sodium acetate or in pyridine.

When refluxed in ethanol containing small amounts of concentrated hydrochloric acid, the above 1,2,4oxadiazoles 4a, b rearrange to the corresponding isoxazoles 2a, b.

The reaction in pyridine has been studied at 35° by using <sup>1</sup>H and <sup>13</sup>C NMR spectroscopy. It has been possible to detect an adduct between the carbinolamine intermediate, resulting from the addition of hydroxylamine to the keto carbonyl group, and the starting thiamide.

During the past few years an increasing interest in 1,2,4oxadiazole derivative: has been developed. Some of these attained enough popularity as antitussive agents and as coronary vasodilators and local anaesthetics.<sup>1</sup>

Although many methods<sup>2</sup> are available for the synthesis of 3.5 - disubstituted - 1.2.4 - oxadiazoles, none is satisfactory for the synthesis of the 3-phenacyl derivatives.<sup>3</sup> We here report the synthesis of the oxime of such derivatives from acetylated or benzoylated benzoylthiacetamide **1a**, **b** and hydroxylamine hydrochloride in ethanol in the presence of sodium acetate or in pyridine.

We also discuss the reaction pathway and, on account of the detailed informations available by the <sup>1</sup>H and <sup>13</sup>C NMR spectra, we present direct evidence of a tetrahedral carbinolamine intermediate, whose existence has been elegantly proved by Cocivera and Effio for nucleophilic additions of NH<sub>2</sub>OH to carbonyl compounds.<sup>4</sup>

Previous studies<sup>3</sup> concerning the cyclization of 1a, b to isoxazoles are reinvestigated for examining the different reactivity of carbonyl and thiocarbonyl groups.

#### **RESULTS AND DISCUSSION**

Synthesis. According to previous results,<sup>5</sup> treatment of **1a**, **b** with hydroxylamine hydrochloride in boiling ethanol led to the formation of the isoxazoles **2a**, **b** (Scheme 1), whose structures were confirmed on the basis of spectral data. Particularly, the presence of the fragments at m/e 105 and 159 in the mass spectrum of **1a** (Table 1) and m/e 105 and 97 in that of **1b**, due to the cleavage of the O-N and C<sub>4</sub>-C<sub>5</sub> bonds, appeared to be diagnostical for the structural assignments. Examination of the crude reaction mixture by <sup>1</sup>H NMR showed that **2** was the only isomer formed.

hydroxylamine hydrochloride in ethanol, in the presence of sodium acetate, or in pyridine for 2 hr at 50°, gave only 4a, b in nearly quantitative yields (Scheme 1). The structures were established by mass spectra, which showed the characteristic fragmentation pattern of the 1,2,4-oxadiazole nucleus,<sup>6</sup> and by NMR spectra. Attempts to obtain phenacyl derivatives, by hydrolyzing the oxime, failed. 1,2,4-Oxadiazoles 4a, b underwent ready rearrangement to isoxazoles 2a, b, by refluxing in ethanol containing small amounts of concentrated hydrochloric acid.

Mechanism. To elucidate the reaction pathway, firstly, <sup>1</sup>H and <sup>13</sup>C NMR parameters were obtained for 1a. In several solvents (pyridine-d<sub>5</sub>, benzene-d<sub>6</sub> and dimethylsulfoxide-d<sub>6</sub>) proton resonances for an intramolecularly H-bonded tautomer 5, whose structure was confirmed by <sup>13</sup>C NMR data, were observed. Transformation into the keto form 1a occurred very quickly in the presence of small amounts of bases. <sup>1</sup>H NMR parameters for both keto and enol forms of 1a are given in Table 2.

The <sup>13</sup>C NMR spectrum of Ia, in pyridine-d<sub>5</sub>, is reported in Fig. 2(a). The <sup>13</sup>C chemical shifts are given in Table 3. Resonances were assigned on the basis of their characteristic chemical shifts<sup>7</sup> as well as the benzoyl-acetamide enol form (Table 3). As follows from the model compound chemical shifts, carbonyl chemical shift differs greatly (19.5 ppm) between keto and enol forms, as observed by Levy.<sup>8</sup>

Acetylated benzoylthiacetamide 1a was then dissolved in pyridine-d<sub>5</sub> containing one equivalent amount of hydroxylamine hydrochloride (1:1 molar ratio) and the resulting mixture was examined by 'H NMR spectrometry shortly after preparation.

Moreover, treatment of la, b with an excess of

Figure 1 illustrates the 90 MHz <sup>1</sup>H NMR spectra, at 35°, obtained before and after (15 min) the addition of

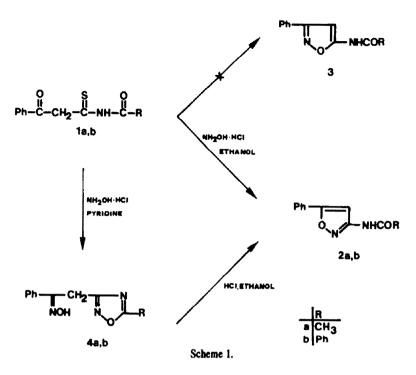


Table 1. Mass Spectra (70 eV) of compounds 2a and 4a

Compound 2a	<u>m/e</u> (rel.intensity)					
	266(5), 265(25), 264(100,M <sup>+</sup> ), 263(21),					
	249(12), 237(5), 236(22), 223(9),222(31),					
	221(8), 220(6), 209(5), 208(7), 207(6),					
	206(5), 187(5), 159(5), 118(5), 106(39),					
	105(51), 102(12), 77(36), 76(8), 63(5),					
	52(5), 51(24).					
4a	218(9), 217(61,M <sup>+</sup> ), 202(11), 201(41),					
	187(23), 175(22), 160(16), 159(40),					
	146(29), 145(55), 128(21), 117(11),					
	105(58), 104(42), 103(100), 98(41),					
	77(69), 60(17), 56(22), 52(16), 51(31),					
	50(13), 43(64).					

### Table 2. <sup>1</sup>H NMR (90 MHz) chemical shifts<sup>a</sup> of la<sup>b</sup>

Solvent	(enol)	(keto) -	-CH(enol)	-CH <sub>2</sub> (keto)	-SH(enol)	-NH(enċi)	-NH(keto)	ArH
с <sub>6</sub> 0 <sub>6</sub>	1.12(=)	-	8.71(s)	-	15.87(bs)	_c	-	7.3-8.3(m
(CD3)250d	2.20(=)	2.13(=)	8.18(=)	5.03(=)	15.73(bs)	11.55(ba)	_c	7.2-8.3(#
C5D5N	2.28(=)*	-	8.78(=)	-	14.33(bs)	12.27(bs) <sup>d</sup>	-	7.3-8.3(#

<sup>a</sup>Chemical shifts in parts per million (ð) from internal Ma<sub>4</sub>Si. Multiplicity:m,multiplet;s,singlet;bs,broad singlet.

Concentrations of 7% w/v were used.

<sup>C</sup>Masked by aromatic protons.

d<sub>Keto/Enol ratio:1/5</sub>.

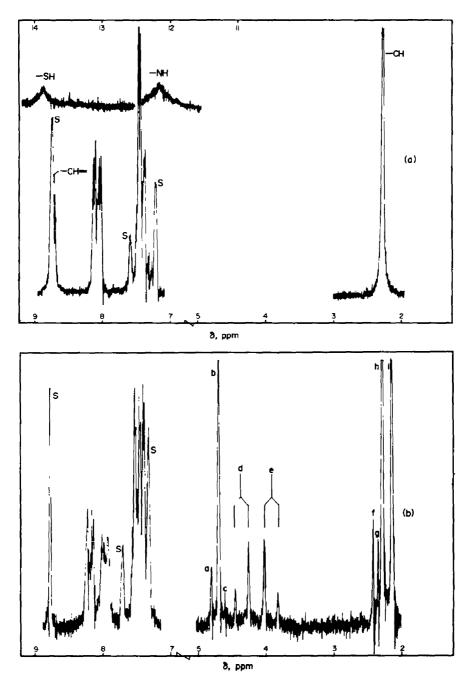


Fig. 1. 90 MHz <sup>1</sup>H NMR spectra, at 35°, of 1a, in pyridine-d<sub>5</sub>, before (a) and 15 min after the addition (b) of one equivalent of NH<sub>2</sub>OH·HCl. The labelled signals are due to the CH<sub>2</sub> and CH<sub>3</sub> protons of monoxime 8 (a, f), adduct 7 (b, d, e, h, i) and final product 4a (c, g).

hydroxylamine hydrochloride to the solution of 1a in pyridine-d<sub>5</sub>.

Surprisingly, several intermediates have a mean life long enough with respect to the NMR time scale. As can be seen from Fig. 1(b), the peaks of the starting material, in the enol form, disappeared immediately and were replaced by an AB quartet (labeled d and e), centered at  $\delta 4.20$  (J = 19.1 Hz), and a singlet at  $\delta 4.74$  (labeled b), in the region of the methylene protons, and two Me signals at  $\delta 2.17$  and  $\delta 2.33$  (labeled h and i). These signals retained an integration ratio 2:2:3:3 during the whole time of the reaction. Such a spectral feature can be ascribed to an adduct between the starting compound in the keto form and the tetrahedral carbinolamine intermediate 6, in which the methylene protons resonate as an AB-type multiplet, being diastereotopic. A tentative structure of such an adduct is represented by 7. No relative configuration of the chiral centers has been attempted.

Little amounts of the 1,2,4-oxadiazole 40 were also observed (singlets at  $\delta 3.72$  and  $\delta 2.41$ , labeled c and g).

Resonances at  $\delta 4.88$  and  $\delta 2.38$  (signals a and f), are attributable to the monoxime 8. In fact, after that the equilibrium was reached with only one equivalent of hydroxylamine hydrochloride at 35°, upon increasing the probe temperature from 35 to 60°, the intensity of the

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Table 3. <sup>13</sup>C NMR chemical shifts in pyridine-d<sub>5</sub>\*

		C-2	C-3	C-4	C-5
с6H5-c=cH-c=-NHCbcH3	177.2	100.7	195.1	169.5	25.0
$c_{6}H_{5} - c_{7}^{1} = c_{1}^{2} - c_{2}^{2} = NH_{2}$	176.3	89.3	170.9		
с <sub>6</sub> н <sub>5</sub> -с <sup>2</sup> -с <sup>2</sup> -хн <sub>2</sub>	195.8	47.6	170.1		
с <sub>6</sub> н - с <sup>3</sup> - с <sup>4</sup> - с <sup>3</sup> - NHCoCH <sub>3</sub>	<u>c</u>	42.2 <sup>b</sup>	195.0	168.1	23.1
с <sub>6</sub> н <sub>5</sub> - с <mark>1-с1</mark> смнсосн <sub>3</sub>	192.8	49.7 <sup>b</sup>	195.0	168.1	23.1
с <sub>6</sub> н - с <sup>1</sup> - с <sup>1</sup> - с <sup>1</sup> - N NOH NO <sup>-</sup> с <sup>1</sup> - N No <sup>-</sup> с <sup>1</sup> - с <sup>1</sup> - N	c	23.4	168.0	177.3	12.0
С <sub>6</sub> H = c <sup>1</sup> -c <sup>4</sup> - с <sup>3</sup> - N NoH N, c <sup>2</sup> - с <sub>6</sub> H 5	<u>_</u>	23.9	169.0	177.1	_
	$c_{6}H_{5} = c_{0}^{1} = c_{0}^{2} - c_{0}^{2} = NH_{2}$ $c_{6}H_{5} = c_{0}^{2} - c_{1}^{2} - c_{0}^{2} - NH_{2}$ $c_{6}H_{5} = c_{0}^{1} - c_{1}^{2} - c_{0}^{2} - NHc^{2} - c_{1}^{2}$ $c_{6}H_{5} = c_{0}^{1} - c_{1}^{2} - c_{0}^{2} - NHc^{2} - c_{1}^{3}$ $c_{6}H_{5} = c_{0}^{1} - c_{1}^{2} - c_{0}^{2} - NHc^{2} - c_{1}^{3}$ $c_{6}H_{5} = c_{0}^{1} - c_{1}^{2} - c_{0}^{2} - NHc^{2} - c_{1}^{3}$ $c_{6}H_{5} = c_{0}^{1} - c_{1}^{2} - c_{0}^{2} - NHc^{2} - c_{1}^{3}$ $c_{6}H_{5} = c_{0}^{1} - c_{1}^{2} - c_{0}^{2} - NHc^{2} - c_{1}^{3}$	$\begin{array}{c} c_{0}H_{5} = c_{1}^{1} = c_{1}^{2} - c_{2}^{2} - NH_{2} \\ c_{6}H_{5} = c_{0}^{1} - c_{1}^{2} - c_{2}^{2} - NH_{2} \\ c_{6}H_{5} = c_{1}^{0} - c_{1}^{2} - c_{2}^{2} - NH_{2} \\ c_{6}H_{5} = c_{1}^{0} - c_{1}^{2} - c_{2}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{2} - c_{2}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{2} - c_{2}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{2} - c_{2}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{2}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{2}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{2}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{3}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{3}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{3}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{3}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{3}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{3}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{3}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{3}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{2}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{1}^{2} - c_{2}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{1}^{1} - c_{1}^{2} - c_{1}^{2} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{1}^{1} - c_{1}^{1} - c_{1}^{1} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - c_{1}^{1} - c_{1}^{1} - c_{1}^{1} - C_{1}^{1} - c_{1}^{1} - NHcocH_{3} \\ c_{6}H_{5} = c_{1}^{1} - $	$\begin{array}{c} c_{0} H_{5} = c_{1}^{1} = c_{1}^{2} - c_{2}^{2} - NH_{2} & d \\ c_{6} H_{5} = c_{0}^{1} - c_{1}^{2} - c_{2}^{2} - NH_{2} & 195.8 & 47.6 \\ \hline c_{6} H_{5} = c_{1}^{0} - c_{1}^{2} - c_{2}^{2} - NHc^{0} c_{1}^{2} & g \\ c_{6} H_{5} = c_{1}^{1} - c_{1}^{2} - c_{2}^{2} - NHc^{0} c_{1}^{2} & g \\ c_{6} H_{5} = c_{1}^{1} - c_{1}^{2} - c_{2}^{2} - NHc^{0} c_{1}^{2} & g \\ c_{6} H_{5} = c_{1}^{1} - c_{1}^{2} - c_{2}^{2} - NHc^{0} c_{1}^{2} & g \\ c_{6} H_{5} = c_{1}^{1} - c_{1}^{2} - c_{2}^{2} - NHc^{0} c_{1}^{2} & g \\ NOH & N_{0} & c_{1}^{2} - c_{1}^{2} & g \\ \hline c_{6} H_{5} = c_{1}^{1} - c_{1}^{2} - c_{2}^{2} - NHc^{0} c_{1}^{2} & g \\ NOH & N_{0} & c_{1}^{2} - c_{1}^{2} & g \\ \hline c_{6} H_{5} = c_{1}^{1} - c_{1}^{2} - c_{2}^{2} - N & c_{1}^{2} & g \\ \hline c_{6} H_{5} = c_{1}^{1} - c_{1}^{1} - c_{2}^{2} - N & c_{1}^{2} & g \\ \hline c_{6} H_{5} = c_{1}^{1} - c_{1}^{1} - c_{2}^{2} - N & c_{1}^{2} & g \\ \hline c_{6} H_{5} = c_{1}^{1} - c_{1}^{1} - c_{2}^{2} - N & c_{1}^{2} & g \\ \hline c_{6} H_{5} = c_{1}^{1} - c_{1}^{1} - c_{2}^{2} - N & c_{1}^{2} & g \\ \hline c_{6} H_{5} = c_{1}^{1} - c_{1}^{1} - c_{1}^{2} - c_{1}^{2} - N & c_{1}^{2} & g \\ \hline c_{6} H_{5} = c_{1}^{1} - c_{1}^{1} - c_{1}^{2} - c_{1}^{2} - N & c_{1}^{2} & g \\ \hline c_{6} H_{5} = c_{1}^{1} - c_{1}^{1} - c_{1}^{2} - c_{1}^{2} - N & c_{1}^{2} & g \\ \hline c_{6} H_{5} = c_{1}^{1} - c_{1}^{1} - c_{1}^{2} - c_{1}^{2} - N & c_{1}^{2} & g \\ \hline c_{6} H_{5} = c_{1}^{1} - c_{1}^{1} - c_{1}^{2} - c_{1}^{2} - N & c_{1}^{2} & g \\ \hline c_{6} H_{5} = c_{1}^{1} - c_{1}^{1} - c_{1}^{2} - c_{1}^{2} - N & c_{1}^{2} & g \\ \hline c_{6} H_{5} = c_{1}^{1} - c_{1}^{1} - c_{1}^{1} - c_{1}^{1} - c_{1}^{2} - c_{1}^{2} & c_{1}^{2} $	$\begin{array}{c} c_{0}H_{5} = c_{0}^{1} = c_{0}^{2} - c_{0}^{2} - NH_{2} & 176.3 & 89.3 & 170.9 \\ c_{6}H_{5} = c_{0}^{2} - c_{1}^{2} - c_{0}^{2} - NH_{2} & 195.8 & 47.6 & 170.1 \\ \hline c_{6}H_{5} = c_{0}^{2} - c_{1}^{2} - c_{0}^{2} - NHCOCH_{3} & \underline{c} & 42.2^{b} & 195.0 \\ c_{6}H_{5} = c_{0}^{1} - c_{1}^{2} - c_{0}^{2} - NHCOCH_{3} & 192.8 & 49.7^{b} & 195.0 \\ \hline c_{6}H_{5} = c_{0}^{1} - c_{1}^{2} - c_{0}^{2} - NHCOCH_{3} & 192.8 & 49.7^{b} & 195.0 \\ \hline c_{6}H_{5} = c_{0}^{1} - c_{1}^{2} - c_{0}^{2} - NHCOCH_{3} & \underline{c} & 23.4 & 168.0 \\ \hline \end{array}$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$

<sup>Ppm</sup> relative to internal TMS; May be interchanged;

"Masked by solvent signals;

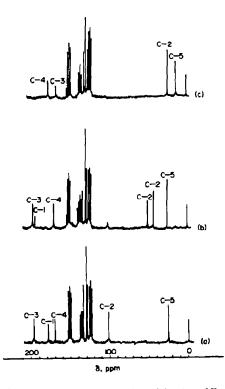


Fig. 2. <sup>13</sup>C NMR spectra (20.1 MHz), in pyridine-d<sub>5</sub>, at 35°, of 1a before (a) and after (b) the addition of one equivalent of NH<sub>2</sub>OH·HCl and of 4c (c). The labelling corresponds to that given in Table 3.

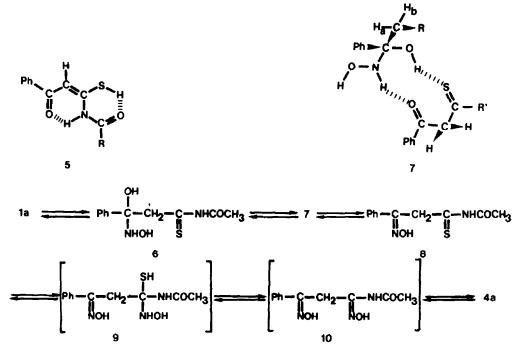
<sup>o</sup>Keto/enol ratio: 2/3.

signals labeled a and f increases at the expense of the adduct 7, the signals due to 4a remaining unchanged. In the alternative assignments of a and f signals to 9 and 10 intermediates, an increase of the cyclization product signals would be expected.

In the presence of an excess of hydroxylamine hydrochloride (1:3 molar ratio), the reaction rate increased and nearly quantitative formation of 4a was observed in 4 hr at 35°. A small amount (5%) of the *anti*-isomer also could be detected.

In order to confirm the initial addition of hydroxylamine to the C=O and the formation of the car-binolamine intermediate, <sup>13</sup>C NMR spectrum of 1a also have been recorded, in pyridine-d<sub>5</sub>, after the addition of one equivalent of hydroxylamine hydrochloride. The resulting spectrum is shown in Fig. 2(b). The signals at 100.7 and 177.2 ppm disappeared and were replaced by two signals at 42.2 and 49.8 ppm, which are triplets in the fully coupled spectrum, attributable to the methylene carbons of the dimeric complex between 1a in the keto form (CO signal at 192.8 ppm) and the carbinolamine intermediate 6. No signals of 4a were present. All <sup>13</sup>C NMR data are summarized in Table 3. Resonances of 4a were assigned on the basis of their characteristic chemical shifts. The signal at 177.3 ppm was assigned to C-4 on the basis of the expected oxygen deshielding, as observed in similar pentatomic heterocycles.<sup>9</sup>

The following conclusions can be drawn from the above results: (1) the C=O group is more reactive, ir pyridine, than the C=S and the addition of hydroxy lamine to C=S follows the dehydration of 6 to produce



Scheme 2.

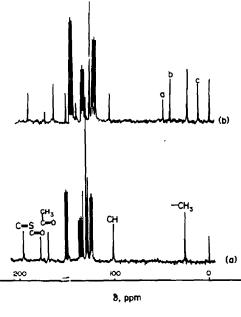


Fig. 3.

the oxime 8, unable to cyclize to the isoxazole 3; (ii) the starting thiamide, by forming a dimeric structure with the carbinolamine intermediate 6, probably plays an important role in promoting hydrogen transfer from N to the O-atom in the dehydration step (several authors attribute this role to water in a number of reaction in such medium<sup>10</sup>); (iii) although no intermediate was observable, by NMR spectroscopy, during the reaction of 1a with hydroxylamine hydrochloride in ethanol, we can hypothesize, on account of the sensitivity of 4a to

the acidic hydrolysis, that the formation of the isoxazole nucleus in such medium, is attributable to a higher reactivity of the thiocarbonyl group in the presence of acid.

#### EXPERIMENTAL

Proton NMR spectra were recorded on a Perkin-Elmer R-32 spectrometer operating at 90 MHz, with TMS as an internal standard. The CMR spectra were recorded as Poulse Fourier transformed NMR (20.1 MHz; Bruker WP-80) spectra with broad band decoupling using deuterium signal lock. Typical conditions were: 5 kHz width; 8 K data; pulse width 1  $\mu$ sec (17<sup>o</sup>). Mass spectra were obtained at 70 eV by direct insertion into the ion source of a LKB 9000S instrument. Elemental analyses were checked with a C. Erba Elemental Analyzer. M.ps are uncorrected.

#### Preparation of the solution for NMR-measurements

150 mg (0.72 mmol) of 1a were dissolved in 1.5 ml of pyridined<sub>5</sub>, containing 1% TMS. 50 mg (0.72 mmol) of  $NH_2OH$ -HCl were then added.

*Materials.* Compounds 1a, b and 2a, b were prepared as described previously,<sup>3</sup> whereas benzoylacetamide was prepared by Haufer method.<sup>11</sup>

#### 3-Phenacyloxime-5-substituted-1,2,4-oxadiazoles 4a, b

(a) To a mixture of 50 mmol  $NH_2OH HCl$  in 100 ml EtOH containing 2 g NaOAc were added 10 mmol of 1a. The mixture was stirred for 2 hr at 50°. The resulting ppt was removed by filtration, washed with water and crystallized from EtOH, yield 85%.

(b) To a soln of 10 mmol of 1a in 25 ml pyridine were added 50 mmol NH<sub>2</sub>OH·HCl. The mixture was stirred for 2 hr at 50°. After pouring into water (200 ml), the resulting ppm was removed by filtration and crystallized from EtOH, yield 80%. M.p.  $151-2^{\circ}$  (Found: C, 60.68; H, 4.58; N, 19.57. Calc. for C<sub>11</sub>H<sub>11</sub>N<sub>3</sub>O<sub>2</sub>: C, 60.81; H, 5.10; N, 19.34%).  $\delta$ (Py-d<sub>5</sub>) 2.28 (3 H, s, CH<sub>3</sub>), 4.58 (2 H, s, CH<sub>2</sub>), 7.0–8.2 (5 H, m, ArH), 14.0 (1 H, broad singlet, NOH). Similar results were obtained for 4b. M.p. 86–7°. (Found: C, 66.85; H, 4.69; N, 15.23. Calc. for C<sub>16</sub>H<sub>13</sub>N<sub>3</sub>O<sub>2</sub>: C, 68.80; H, 4.69; N, 15.05%).  $\delta$ (Py-d<sub>5</sub>) 4.76 (2 H, s, CH<sub>2</sub>), 7.3–8.4 (10 H, m, ArH), 14.0 (1 H, broad singlet, NOH).

Hydrolysis of 4a, b to 2a, b. A soln of 10 mmol of 4a, b in 20 ml EtOH and 1 ml conc HCl was refluxed for 1 hr. The mixture was cooled to room temp and the resulting white crystals were filtered off and crystallized from EtOH. These products were identical in all respects with 2a, b, prepared from 1a, b with hydroxylamine hydrochloride in EtOH.<sup>5</sup>

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#### REFERENCES

- <sup>1</sup>F. Eloy and R. Lenaers, Bull. Chim. Therap. 347 (1966); M. DeGregorio, Panminerva Med. 4, 90 (1962); K. Harsanyi, P. Kiss, D. Kobonits and I. R. Malyata, Arzmeim.-Forsch. 16, 615 (1966); J. Sterne and C. Hirsch, Therapie 20, 89, 95 (1965).
- <sup>2</sup>A. R. Katritzky and A. J. Boulton, Advances in Heterocyclic Chemistry, pp. 65-78. Academic Press, New York (1979).
- <sup>3</sup>R. Merckx, Bull. Soc. Chim. Belges 56, 339 (1947); 58, 58 (1949).

- <sup>4</sup>M. Cocivera and A. Effio, J. Chem. Soc. Chem. Comm., 393 (1976); M. Cocivera and A. Effio, J. Am. Chem. Soc. 98, 7371 (1976).
- <sup>5</sup>B. Tornetta, Ann. Chim. Italy 53, 244, 253 (1963).
- <sup>6</sup>A. Selva, L. F. Zerilli, B. Cavalleri and G. C. Gallo, Org. Mass Spectrom. 6, 1374 (1972); 9, 558 (1974).
- <sup>7</sup>N. N. Shapet'ko, S. S. Berstova, G. M. Lukovkin and Yu. S. Bogachev, Org. Magn. Res. 7, 239 (1975); C. Piccinni-Leopardi, O. Fabre, D. Zimmermann, J. Reisse, F. Cornea and C. Fulea, *Ibid.* 8, 536 (1976).
- <sup>8</sup>G. C. Levy and G. L. Nelson, *Carbon-13 Nuclear Magnetic Resonance for Organic Chemists*. Wiley-Interscience, New York (1973).
- <sup>9</sup>H. Hiemstra, H. A. Houwing, O. Possel and A. M. vanLeusen, *Can. J. Chem.* 57, 3168 (1979); J. Gainer, G. A. Howarth and W. Hoyle, *Org. Magn. Res.* 8, 226 (1976).
- <sup>10</sup>J. E. Riemann and W. P. Jenks, J. Am. Chem. Soc. 88, 3973 (1966).
- <sup>11</sup>C. R. Haufer and C. J. Eby, *Ibid.* 79, 725 (1957).